# Cost Effective Bioinformatics Pipeline for NGS Reporting of Clinically Relevant Cancer Genes Using the Ampliseq Cancer Hotspot Panel v2

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### **BACKGROUND**

Laboratories with limited resources may lack an adequate bioinformatics pipeline to analyze and report NGS results. Our study aimed to develop an affordable solution for reviewing clinical NGS data.

#### **MATERIALS AND METHODS**

Variant caller (VCF) files were generated by the Variant Caller (VC) plug-in provided with the Ion PGM sequencer (high stringency) and 3rd party software. NextGene (NG) by Softgenetics (low stringency). VCF files, patient demographics, and NGS QC metrics were transferred to software developed in Excel Visual Basic, Analysis consists of two automated macros, prompted by user input. The first script checks rolling databases to annotate and sort raw data based on various filters, including mutation frequency (<1%), coverage (<500x), known SNPs, and sequencing artifacts. VCF files from VC and NG are compared and common variants are identified. A manual review of nucleotide sequences is performed for a predefined subset of genes based on tumor type. Technologists evaluate the annotated data and verify the mutations to report. The second script then compiles a comprehensive report based on mutational findings and patient demographics. To ensure technical accuracy, 131 known positive cases were analyzed using the bioinformatics pipeline and clinically relevant findings were validated via Sanger Sequencing. Cases consisted of pancreatic neoplasm (34%), lung carcinoma (15%), tumor cell line (15%), desmoids tumors (10%), myeloid neoplasm (9%), melanoma (7%) colon carcinoma (4%) and GIST (3%).

#### **NGS Bioinformatics Workflow**

Variant Caller Raw Data VCF is obtained from Variant Caller (high stringency) Nextgene Raw Data
VCF is obtained from NextGene
(low stringency)

Excel NGS Analysis Software Raw data is imported into Excel NGS Analysis Software.

Tumor Type	Pancreatic Cyst	¥			
% Tumor	Colon Cancer (CRC) Breast Cancer	^			
Mutation Frequency Threshold	Lung Cancer				
	Pancreatic Cyst				
Additional Information for AML	Malignant Melanoma Thyroid Cancer				
Baseline?	GIST	~			

## **NGS Bioinformatics Workflow (Continued)**

#### **Pre-Review**

Software checks rolling databases to annotate and sort raw data

Figure 1: Excerpt of Filtered VC and NG Raw Data from Excel Analysis Software.

DMO Number Analysis By Fight Stringency							DMO Number	Analysis By	J					L	ow	string	gency						
MD15-XXX RS >2% frequency, >100x coverage							MD15-XXX	RS.									uency, >	300	)x cov				
Beview		Chrom	Position	Chr.Positio	Ref	Variant	Frequency	Quality	Allele Nan	Gene ID	Review	Index	Gene	r.,	Chr.Position	nafe.	C-J				Mutation Cal		
<mut i<="" td=""><td></td><td>chr20</td><td>57484421</td><td>20 574844</td><td>G</td><td>4</td><td>3.8</td><td>20.508</td><td></td><td>9GNAS</td><td>dMUIT FRED TH</td><td>IIIUOX 104</td><td>CHAL</td><td>VIII.</td><td>20 57494421</td><td>A COL</td><td>C C</td><td>OVE AGE</td><td>SUITE</td><td>3/8" GD</td><td>WORDS COLC</td><td>3.36</td><td>MILLIO ALIO</td></mut>		chr20	57484421	20 574844	G	4	3.8	20.508		9GNAS	dMUIT FRED TH	IIIUOX 104	CHAL	VIII.	20 57494421	A COL	C C	OVE AGE	SUITE	3/8" GD	WORDS COLC	3.36	MILLIO ALIO
		chr12	25398284	12.253982		ă.	12.7		COSM520		CHIQI PRZQ IP	200	KRAS	11	12 25398284	,	0	2401	26.6	** 1310	c.35G>TG	12.0	12G×VG
		chr7		7.1163396		Ť	51.4		COSM706		_		MFT		7 116339642		0 (	2625			c.504G>GT		168E>0E
QUP.CI	IIFNT	chr10		10.436138			100	31914		RET			SMAD4		18.48604733	9	Δ.	2209	15.2		c.1557 1558		
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SNP-M	ISSENS	chr4	55972974	4.5597297		A	49.1	9875.2	-	KDR	1		GNA11		19.3118934	ċ	c	671	1.0	120707.	c.623 624in		
SNP-M	ISSENS	chr17	7579472	17,757947	G	Ċ	100	31736.	3	TP53	SMP-SILENT		APC .		5.112175770	6	G	2740	25.8	rs41111	9 6479654		1493T>T
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INTRO	N	chr22	24176287	22,241762	G	A	49.7	9792.6	COSM109	SMARCB1	CMP-MISCENCE	35	KDR	-	4 55977974	-	4	1937	23.8	rs1870	14164oTA	49.9	4720>H0
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							INTRON	48	NPM1		5.170837530	Ġ	G	1128	6.5		IVS847-2 84	2.22:1	Solice				
							Homopolymes	101	STK11	19	19.1207065	Ġ	G	4021	9.3		c.157 158ire	6.74:2	FS				
								Homopolymes	102	STK11	19	19.1207077	Ġ	G	4018	10.9		c.169 170 in	6.05:2	FS			
Blue: Sequencing Artifact Red: Less than 5% mutation frequency threshold Orange: Less than 500X coverage							Homopolymes	84	IDH2	15	15,50631918	¢	G	2055	3.6		c.435 436ins	8.52;1	FS				
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							Deletion Artifi	60	FGFR1		8,38285916	A	T	1179	21.8			1.87					
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Pink: Less than 500X coverage and 5% mutation frequency threshold								Artifact	95	TP53	13	17,7579473	Ġ	C	2086	13.8		t.214C>6C	9.16	72P>AP			
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٠											<mut freo="" td="" th<=""><td></td><td>SMARCR1</td><td></td><td>22.24143265</td><td></td><td></td><td>1374</td><td></td><td></td><td>c.499 500ire</td><td>182</td><td></td></mut>		SMARCR1		22.24143265			1374			c.499 500ire	182	

#### Mutation Review

Technologist review of sequence traces to confirm bio-informatic calls (Cells in blue above highlight variants common to NG and VC)

#### Post-Review

Software automatically generates a comprehensive final report from mutational findings and patient demographics.

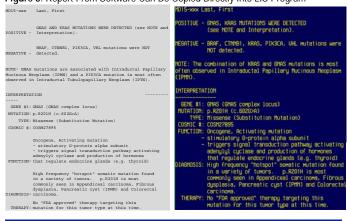
Figure 2: Excerpt of Final Report Generated by Excel Analysis Software

Ion Torrent N	ext Generation	on Se	equencing Re	port		QC Review	PRE-REVI					
						Date						
Patient Name	Last, First	1	Analysis By	Rebecca				POST-REVI				
DMO#	MD15-XXX	1	Analysis Date	10/28/2015		Excel Macro Version	10122015 v1	1 OST ILEVI				
Cell Block/Thin Prep#	na	1	PGM Run #	500		Variant Caller Reference	hg19 tmap-f3					
Patient Sex	Female	1	Barcode #	IonXpress 003		Nextgene Reference	Human v37 3 dbsnp135 dna	MEDITEC				
Patient Date of Birth	10/28/1920		PGM Instrument #	PGM2		COSMIC version	v72	IVILDITEC				
Patient Age	95		Tumor Type	Pancreatic Cyst		dbSNP version	Build 127					
Medical Record #	123456789		Tumor %	10-20%				MEDITEC				
Chip ID #	10-11-15-1		Analysis Threshold									
Control Name	Supercontrol		AML Baseline?	NA				FAILURE				
Control Lot #	011415		Transplant Status	NA								
Detected Mutat												
Common Findings:	GNAS (201R>HR), KR	RAS (126	5>VG), MET (168E>DE	) mutations were detected	by Variant Calle	r and Nextgene.						
Variant Caller:	See common finding	s.										
Nextgene:	See common finding	s.										
"See next page for detailed r	mutation report.											
				MODIFY NEX	TGENE							
Manual review	of clinically rel	evant	t genes	WOODII T INEX	TOLIVE	J						
	Chromosome	T T	ľ		Wildtype or							
Gene	location of ROI	Exon	Codons covered	Coverage (depth) x	Mutant?	1 1	Place Patient Barcode Here (From LIS system)					
BRAF	7,140481403	11	469	793	Wildtype							
BRAF	7,140453136	15	598-601	1179	Wildtype							
CDKN2A	9,21971185	2	58	5413	Wildtype							
CDKN2A	9.21971120	2	80	5391	Wildtype	í l						
GNAS	20.57484421	8	201	878	MUTANT	[ L						
GNAS	20.57484596	9	227	2720	Wildtype		Sample ID HID Kev	F-TRYASAW				
KRAS	12.25398285	2	12-13	2655	MUTANT							

## **NGS Bioinformatics Workflow (Continued)**

#### **Laboratory Information System (LIS) Report**

Software automatically compiles formatted LIS Reports
Figure 3: Report From Software Can Be Copied Directly Into LIS Program



#### **RESULTS**

- •All clinically relevant mutations (100%) were detected by the NGS bioinformatics pipeline and by Sanger Sequencing.
- On average, VC made 22 calls per case, whereas NG made 86 calls.
   Of the total variants per case, an average of 24% were identified as common SNPs, 8% as sequencing artifacts, 42% as low frequency artifacts (<1% allele frequency), and 2% as variants with low coverage</li>
- An average of 3 variants per case was common to both VC and NG.

(<500x), ultimately filtering approximately 77% of the raw data.

•On average 2 somatic mutations per case (67%) and 1 rare hereditary germline variant (33%) were identified, accounting for 9% of total VC findings and 2% of total NG findings.

#### **DISCUSSION/CONCLUSION**

- •This pipeline is an affordable and accessible alternative to commercially available products or bioinformatics team.
- •The combination of two software packages into one pipeline reduces the likelihood of error and ensures accurate reporting.
- •Automation of processes such as data sorting and report compilation reduces resources needed to analyze clinical samples.